Diastereoselective functionalizations of enecarbamates derived from pipecolic acid towards 5-guanidinopipecolates as arginine mimetics[†]

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Various substituents could be diastereoselectively introduced into the 5-position of pipecolic acid *via* electrophilic or free-radical-initiated addition to the carbon–carbon double bond of endocyclic enecarbamates derived from pipecolic acid. This study allowed the diastereoselective synthesis of both *cis*- and *trans*-5-guanidino pipecolates, which were designed as constrained arginine mimetics and whose potential inhibition of nitric oxide synthase (NOS) was evaluated with three NOS isoforms.

Introduction

Pipecolic acid is an important noncoded cyclic amino acid, which is found in numerous microbial, plant and animal species, including humans; it is an intermediate in the major path of Llysine degradation in the central nervous system.¹ Free L-pipecolic acid and many of its derivatives are found in nature.² In recent years it has been incorporated into several peptidic structures as a proline homologue.³ In addition, some derivatives of substituted pipecolic acid were developed as β-turn mimetics,⁴ and others were designed as constrained analogues of lysine⁵ or phenylalanine,⁶ or as N-methyl-D-aspartate receptor antagonists.7 We have been working for some years on the chemistry of enecarbamates,⁸ especially those derived from pipecolic acid.9 As part of our study in this field, we required efficient access to 5-guanidinopipecolates 17 (Scheme 1), which we designed as arginine mimetics for their potential as NO synthase (NOS) inhibitors.10 In connection with this project, we report here the results of our complete study on various regio- and stereoselective functionalizations of endocyclic enecarbamates 3, in order to develop efficient access to both diastereomers of the target molecules 17 (Scheme 1).

Although there have been some studies describing reactions of enecarbamates with various electrophiles,¹¹ only a few have involved enecarbamates derived from pipecolic acid.¹² In this study, we used various reaction conditions that enabled us to prepare new pipecolate derivatives efficiently and diastereoselectively. The methodology employed was based on the reaction of 5,6-dehydropipecolate derivatives **3** with various electrophilic and radical species that led to selective functionalization at the C-5 position.



Scheme 1 5-Guanidinopipecolates from enecarbamates 3.

Results and discussion

Substrate preparation

The starting materials, (*i.e.* racemic enecarbamates **3a-c**), were easily prepared from (\pm) -pipecolic acid on a multigram scale according to known procedures (Scheme 2).13 The N-protected methylpipecolates 1a,b were electrochemically oxidized into 2, which underwent subsequent acid-catalyzed methanol elimination producing the corresponding compounds 3a and 3b. We have previously reported that addition of π -nucleophiles onto the iminiums derived from 2a and 2c yielded opposite diastereomers.¹⁴ Consequently, we thought that due to its intrinsic strain, bicyclic enecarbamate 3c might provide different diastereoselectivity from that expected with monocyclic substrates 3a,b. Moderate yields of the bicyclic substrate 3c were obtained by reductive cyclization of 3a.¹⁵ In order to improve the yield of 3c, we investigated a different order of the oxidation-elimination-cyclization sequence used, by carrying out the oxazolidine ring formation prior to electromethoxylation and subsequent elimination of methanol (Scheme 2). Unfortunately, electromethoxylation of oxazolidinone 1c selectively yielded the regioisomer 2'c where the methoxy group was introduced at the ring junction position, as we have already observed with an analogous oxazolidinone derived from proline.16

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Scheme 2 Preparation of enecarbamates 3 from N-protected methylpipecolates.

Addition reactions of enecarbamates 3

5-Azido-6-methoxylation. We devised the synthesis of the desired arginine analogues 17 by guanylation of the corresponding ornithine analogues, *i.e.* 5-aminopipecolates 15, whose synthesis could be accomplished from enecarbamates 3 (Scheme 1). We first explored the β -hydroamination of the enecarbamates **3a** by reacting its crude hydroboration adduct with hydroxylamine-O-sulfonic acid,¹⁷ which failed to provide the desired 5-aminopipecolate derivatives. We then planned to obtain the latter via the corresponding 5-azidopipecolates. To this end, we first examined direct regioselective introduction of this 5-azido substituent by oxidative-radical β -azido- α -methoxylation of the enecarbamate moiety. Preliminary attempts to perform this addition by electrochemical oxidation in a divided cell (Pt-Pt) according to a procedure described by Fujimoto et al.18 led to low yields of the addition compounds 4.¹⁹ We then examined the conditions employed by Chavan et al.,²⁰ who performed azidomethoxylation of electron-rich olefins (including enol ethers), and more recently performed by Norton Matos et al.21 with pyrrolidine and piperidine enecarbamates, by using cerium ammonium nitrate (CAN) as oxidizing agent, instead of anodic oxidation. Treatment of substrates **3a**,**b** with a combination of CAN/NaN₃ in the presence of methanol as the solvent thus led to the corresponding 5-azido-6-methoxypipecolates 4a,b in good yields, albeit as mixtures of three diastereomers according to gas chromatography analysis (Scheme 3). Subsequent treatment of these mixtures of isomers with Et₃SiH and BF₃·OEt₂ at low temperature resulted in the chemoselective reduction of the a-aminoether moieties, thus leading to the corresponding azido derivatives 5a,b as mixtures of epimers at the C-5 position. The *trans/cis* ratio resulting from this sequence was highly dependent on the temperature of the first step, thus varying from approximately 1 : 1 at 0 °C to a 92 : 8 trans/cis mixture at -95 °C in acetone instead of acetonitrile as solvent for the azidomethoxylation step. Bicyclic substrate 3c was less reactive towards azidomethoxylation, unlike **3a,b**, which reacted even at $-95 \,^{\circ}C(3-5h at 1 \text{ mmol scale})$, the total conversion of substrate 3c (1 mmol) required 16 h at room temperature. Under these conditions, 3c led to 4c as a 3 : 7 mixture of epimers at the C-6 position (oxazolidinone numbering). In both isomers the 5methoxy group is cis to the hydrogen atom of the bicyclic junction (8*a*-H of the oxazolidinone **4c**), and is *trans* to the 6-azido group in the predominant epimer. Reductive treatment of this mixture with a Et₃SiH/BF₃·OEt₂ combination led to the same 3 : 7 ratio of *trans/cis* isomers of 5c (Scheme 3). Despite this interesting reverse selectivity that favours the *cis*- over the *trans*-isomer in the case of bicyclic enecarbamate 3c, this method would require at least three steps to reach the cis-5-azido pipecolate from 5c. In conclusion, the azido-methoxylation-reduction sequence seems suitable for the synthesis of trans-5-azidopipecolate when the first step is performed at a low temperature with monocyclic substrates **3a.b** (Scheme 3).

In order to obtain selectively the *cis*-5-azido pipecolate, we devised a two-step process in which the 5-azido substituent could be introduced by nucleophilic displacement of a suitable leaving group having a relative *trans* orientation to the C-2 substituent. Two types of leaving group, *i.e.* halides and sulfonic esters, were explored for this purpose.



Scheme 3 5-Azido pipecolate derivatives from 3.

5-Halo-6-methoxylation. Treatment of enecarbamates **3a,b** with NBS/MeOH or NIS/MeOH at -70 °C in THF afforded good yields of the corresponding *O*-methyl halohydrines **6** with similar diastereoselectivity to that observed in the case of the azidomethoxylation reaction (Scheme 4). Under the same reaction conditions (*i.e.* NIS/MeOH) **3c** led to a single diastereomer of **6'c**, as shown in Scheme 4. Treatment of **6a** with triethylsilane in presence of boron trifluoride diethyl etherate at a low temperature afforded good yields of the demethoxylated derivative **7a** chemoselectively and with high diastereoselectivity (*trans/cis* = 96/4). Under the same conditions, 5-iodo-6-methoxy derivative **6'a** led to a 65 : 35 mixture of the expected 5-iodopipecolate **7'a** and compound **1a**. The formation of the latter was explained by the easy reduction of the carbon–iodine bond under the conditions required for the *N*, *O*-acetal moiety reduction (Scheme 4).



Scheme 4 Halomethoxylation of enecarbamates 3.

Before discussing the halide substitution by azide, we describe the results for the preparation of 5-hydroxypipecolates, which need to be activated as sulfonates prior to substitution with azide *via* $S_N 2$ displacement.

5-Hydroxypipecolate derivatives. Starting from substrates **3**, we explored three series of reactions aiming at the introduction of a 5-hydroxy substituent on the pipecolate ring, *i.e.* oxidative hydroboration, 5,6-dihydroxylation or 5-hydroxy-6-methoxylation followed by acetylation and subsequent chemoselective N,O-acetal reduction.

Oxidative hydroboration of 3. The best results were obtained with $BH_3 \cdot SMe_2$ complex (Scheme 5). Generally, the hydroboration of enecarbamates 3 does not reach the trialkyl stage,²² and moreover, this step is very slow in THF (15 h on a 1 mmol scale). Studying the effects of various solvents showed that improved rates were observed when the hydroboration step was performed in less basic solvents such as diethyl ether, toluene or, in particular, dichloromethane. Moreover, the best yields were obtained when the oxidative cleavage was achieved by removing all volatiles under vacuum, prior to oxidative treatment with trimethylamine *N*oxide (TMO) in refluxing THF.^{23a} Both monocyclic enecarbamates **3a,b** gave the corresponding alcohols *trans*-**8a,b**^{12c,23b} selectively (Table 1, entries 1–5). Lower yields and almost no diastereoselectivity occurred with substrate **3c** (Table 1, entries 6–8).



Scheme 5 Oxidative hydroboration of 3.

 Table 1
 Diastereoselectivity of oxidative hydroboration of 3

Entry	Substrate	Solvent	Alcohol (%)	trans/cis
1	3a	THF	8a (54)	84:16
2	3a	Et ₂ O	8a (45)	83:17
3	3a	CH ₂ Cl ₂	8a (63)	85:15
4	3a	PhMe	8a (63)	75:25
5	3b	CH ₂ Cl ₂	8b (63)	93:7
6	3c	Et ₂ O	8c $(35)^a$	48:52
7	3c	CH ₂ Cl ₂	8c $(20)^a$	45:55
8	3c	PhMe	8c (35) ^a	55:45

^a Up to 35% of compound 1c was obtained.²³

5,6-Dihydroxylation and 5,6-hydroxymethoxylation of 3. Besides the oxidative hydroboration method, we examined two other oxidation methods based on dihydroxylation or hydroxymethoxylation (Scheme 6). Dihydroxylation of substrates 3a,b by using $OsO_4/TMO^{12b,24}$ or $Oxone^{25,12a}$ in an acetone–water mixture led to the expected 5,6-dihydroxypipecolates 9^{24c} mainly as mixtures of two diastereomers. Attempts to selectively reduce the hemiaminal moieties selectively by using a Et₃SiH/BF₃·OEt₂ combination resulted in poor yields of the expected 5-hydroxypipecolates 8, along with the corresponding 5-oxopipecolates 12. The latter were generated via the iminium intermediate which, instead of reduction by Et₃SiH, may also undergo deprotonation leading to the corresponding β -hydroxy enecarbamate which, in turn, tautomerizes into the corresponding 5-oxopipecolate 12. To avoid such rearrangement, crude diols 9 were converted to diacetates 9,^{12c} prior to N,O-acetal reduction with Et_3SiH/BF_3 ·OEt₂ in dichloromethane (Scheme 6). By using such a sequence, we were able to isolate monoacetates $11a, b^{12c}$ in moderate to good overall yields (44-71%), and good selectivity in favour of the trans isomers, whatever the dihydroxylation reagent (Table 2, entries 1-4). Higher overall yields were obtained in the case of substrate 3c; however, the diastereoselectivity depended on the dihydroxylation reagent (Table 2, entries 5 and 6). Thus, the OsO₄/TMO combination led to the trans isomer, although with lower selectivity than that reached with monocyclic substrates, while reverse stereoselectivity was observed when Oxone was employed as dihydroxylating agent. Workup of the vicinal diols 9 was generally difficult, because of their significant hydrosolubility. To avoid these difficulties, substrate 3a was allowed to react with Oxone in methanol, thus producing the expected 5-hydroxy-6-methoxypipecolate derivative 10a, which after acetylation and subsequent reduction afforded good overall yield of 11a, albeit with almost no diastereoselectivity (Table 2, entry 7). When applied to 3c (entry 8), the same three-step sequence led to **11c** with reverse selectivity (*trans/cis* = 3/7) as in the case of azidomethoxylation. It is of note that, when oxidized with Oxone, 3c led to bicyclic compounds (9c or 10c) whereas



Scheme 6 Monoacetates 11 from 3.

 Table 2
 Formation of monoacetates 11a-c from 3a-c

Entry	3	Oxidation conditions	11 (%) ^a	trans/cis
1	3a	OsO ₄ /TMO, H ₂ O	11a (69)	92:8
2	3a	Oxone/H ₂ O	11a (55)	85:15
3	3b	$O_{s}O_{4}/TMO, H_{2}O$	11b (71)	92:8
4	3b	Oxone/H ₂ O	11b (44)	86:14
5	3c	$O_{s}O_{4}/TMO, H_{2}O$	11c (60)	70:30
6	3c	Oxone/H ₂ O	11c (75)	37:63
7	3a	Oxone/MeOH	11a (58)	55:45
8	3c	Oxone/MeOH	11c (84)	30:70

^{*a*} Overall yields starting from **3**.

the C-5 substituents were always *syn* to the hydrogen atom of the bicyclic junction (8a-H), whatever the C-6 relative configuration. *m*-Chloroperbenzoic acid in methanol²⁶ was also employed as oxidizing agent with substrates **3c**; the expected hydroxymethoxyp-ipecolate **10c** was obtained in moderate yield (30%), along with its corresponding 5-hydroxy-6-(*m*-chlorobenzoyloxy)pipecolate. The latter was isolated in 46% yield when the reaction was performed in absence of methanol and sodium bicarbonate.

Diastereoselectivity of the above additions. Some general comments regarding the observed diastereoselectivities in the additions performed on substrates **3** are necessary, before describing the functional conversion required at the C-5 position to synthesize the targeted 5-guanidinopipecolates. Some of the additions described above took place in a rather concerted manner (hydroboration and *syn*-dihydroxylation with osmium tetraoxide), and others (azidomethoxylation, halomethoxylation dihydroxylation or hydroxymethoxylation) involved multistep mechanisms. Generally, when performed on monocyclic substrates **3a,b**, all these additions and subsequent C-6 reduction provided selectively a *trans* relative orientation between the C-2 and C-5 substituents in the

corresponding final monocyclic products (5, 7, 8 and 11), regardless of the mechanism involved in the addition step. However, additions to bicyclic enecarbamate 3c appeared to be mechanismdependent; syn-dihydroxylation with the OsO4/TMO combination provided selectively the trans isomer (addition anti to the oxazolidinone methylene), while syn-hydroboration led to an approximately equimolar cis/trans mixture of alcohol 8c. Other additions, which were multistep reactions, led selectively to final compounds (5c. 7c and 11c) in which the C-6 substituent was *trans* to the hydrogen atom of the ring junction (i.e. 8a-H). To explain the observed diastereoselectivities, we assume that, due to the partial double bond character of their exocyclic carbon-nitrogen bond, monocyclic enecarbamates 3a,b preferentially adopt the envelopelike conformation **IIa.b**, owing to the 1,3-allylic strain present in conformation Ia, b where the 2-carbomethoxy substituent occupies a pseudo-equatorial position²⁷ (Scheme 7). Considering this conformational preference, two factors may account for the facial selectivity: (i) steric repulsion between the adding (electrophile or free radical) species and the 2-carbomethoxy group, and (ii) addition on the α -face (path a, Scheme 7) is expected to proceed *via* a half-chair transition state, while addition on the β -face (path b, Scheme 7) would proceed through a boat-like transition state. Then, for stereoelectronic reasons, both diastereomeric iminiums A and B would add nucleophiles (MeOH or H₂O) mainly syn to the 2-carbomethoxy substituent.²⁸



Scheme 7 Possible explanation of the observed diastereoselectivity with **3a,b**.

Due to its bicyclic structure and resonance, enecarbamate **3c** is locked in conformation **Ic**. The first stage of stepwise additions takes place following path b (Scheme 8) preferentially *via* a half chair-like transition state leading to iminium ion **D**. For stereoelectronic reasons, addition of nucleophiles to both iminium intermediates **C** and **D** takes place exclusively on the α -face¹⁴ (Scheme 8), thus leading to a mixture of epimers at the C-6-position (tetrahydrooxazolopyridin numbering). In both mono- and bicyclic enecarbamates, and unlike stepwise additions,



Scheme 8 Possible explanation of the observed diastereoselectivity with 3c.

syn-dihydroxylation catalyzed with OsO_4 takes place selectively on the α -face, probably due to its concerted character and steric demand of the dihydroxylating osmium complex. However, both diasteromeric syn-diols derived from bicyclic substrate **3c** evolve into a mixture of C-6 epimers, due to epimerization at position C-5, which leads to the favoured *trans*-2,5 relative stereochemistry. Non-stereoselective syn-hydroboration of **3c** could be interpreted as a balance between steric and stereoelectronic factors.

5-Azidopipecolates via $S_N 2$ reactions

Most of the oxidative methods examined above led selectively to the *trans* diastereomer: to obtain the *cis* 5-hydroxypipecolate selectively we decided to examine the reduction of its corresponding ketone 12. This reduction was undertaken by assuming that ketones 12a,b will be locked in a conformation with the 2-carbomethoxy substituent being axially oriented due, to the A^{1,3} allylic interaction usually present in the α -substituted Nacylpiperidines.²⁷ Therefore, due to stereoelectronic control, treatment of such compounds with non-bulky hydride was expected to lead stereoselectively to the cis isomer of alcohol 8, via an axial addition of hydride. Ketones 12^{29} were first obtained by oxidation of alcohols 8; however, equivalent or slightly better yields were obtained following a one-pot procedure (Scheme 9) consisting of hydroboration of enecarbamates 3 and subsequent oxidative treatment with oxidants stronger than TMO or hydrogen peroxide (Table 3, entries 1-4). The best yields were obtained with



a: R¹ = OMe, R² = CO₂Me; **b**: R¹ = OBu^t, R² = CO₂Me; **c**: R¹—R² = -O-CH₂-

Scheme 9 Preparation of cis alcohols 8a,b via ketones 12.

Table 3 One-pot oxidation of 3a-c to ketones 12

Entry	Substrate	Conditions	Ketone (%)
$\frac{1}{2}$	3a 3a 3b 3c	1) BH ₃ ·SMe ₂ , 2) PCC 1) BH ₃ ·SMe ₂ , 2) IBX 1) BH ₃ ·SMe ₂ , 2) IBX 1) BH ₃ ·SMe ₂ , 2) IBX	12a (45) 12a (51) 12b (40) 12c (66)
5	3c	1) <i>m</i> -CPBA, PhMe; 2) PTSA (reflux)	12c (30)

2-iodoxybenzoic acid (IBX) as oxidant in refluxing acetonitrile. A lower yield of ketone **12c** was obtained *via* epoxydation of **3c** with *m*-CPBA and subsequent acid-catalyzed rearrangement (entry 5) as described by Matsumura *et al.*^{29a} Reduction of ketones **12a,b** with sodium borohydride yielded the corresponding *cis* alcohols **8a,b** with high diastereoselectivity (Scheme 9).

As mentioned above, azidomethoxylation of enecarbamates 3a,b allowed selective formation of *trans*-5-azidopipecolates. To synthesise the *cis* isomers selectively, we considered nucleophilic displacement of a leaving group from either trans-5-halogeno- or trans-5-hydroxypipecolates with azide. Attempts to perform such nucleophilic substitution by reacting 5-halo-6-methoxy derivatives 6 (or 6') with sodium azide at 70 °C in DMF led to only partial elimination, which took place selectively at the C-4-C-5 position. Higher conversions were observed when substrates 6 were reacted with DBU or DABCO instead of sodium azide; this regioselective elimination was consistent with the trans-relationship between the 5-halogeno- and 6-methoxy substituents in derivatives 6. This absence of nucleophilic substitution may be ascribed to the steric effect of the 6-methoxy substituent, which prevented the azide ion from reaching the electrophilic C-5 center. We therefore reacted demethoxylated compound 7a with sodium azide at 70 °C in DMF, and obtained a 65:35 mixture of $S_N 2/E2$ compounds. As anticipated, the elimination reaction with substrate 7a was no longer regioselective. When using excess TMSN₃ in the presence of TBAF,³⁰ mainly nucleophilic substitution occurred (Scheme 10). A similar mixture was obtained from alcohol *trans*-8a, by using Mitsunobu conditions³¹ (Scheme 10), albeit with low yields (<29%). Unfortunately, the major azide, *cis*-5a, was difficult to isolate from these mixtures in all cases.



Scheme 10 Azide *cis*-5a from *trans*-7a and *trans*-8a.

Finally, we considered a two-step path, starting from alcohols **8a,b**, which were first activated as sulfonate esters, prior to reaction with sodium azide in DMF (Scheme 11). Indeed, mesylates **13** or tosylates **14** were efficiently converted to the corresponding azides *via* a clean $S_N 2$ process. In addition, both diastereoisomers of alcohols **8a,b** could be successfully involved in this sequence. Thus, while *trans*-5-azides **5a,b** could be obtained either from the corresponding enecarbamates **3a,b** or from alcohol *cis*-**8a,b**, the *cis*



Scheme 11 Azides 5 from alcohols 8.

isomers could only be reached *via* alcohols *trans***-8a,b**. Eventually, both diastereoisomeric 5-azidopipecolates were prepared in pure form and with good yields.

5-Guanidinopipecolates

To reach the desired 5-guanidinopipecolates, each diastereomer of azide 5 had to be converted into the corresponding amine and then guanylated before final deprotection (Scheme 12). Azides 5 were submitted to hydrogenolysis in the presence of 10%palladium-on-charcoal in dichloromethane, and led efficiently to the corresponding primary amines 15, which can be considered as ornithine analogues. To avoid intra- or inter-molecular amidation, amines 15 must be either stored as hydrochloride salts, or employed immediately in the following guanylation step.³² To perform the latter, we tried two guanylation procedures³³ as shown in Table 4. The best yields of guanidine derivatives 16 were obtained by using N, N'-bisprotected-S-methylthiourea in the presence of triethylamine and mercury(II) chloride. We also performed the same hydrogenolysis/guanylation sequence starting from free acids 5'. Isomer trans-16'b was obtained with 58% yield, while guanylation of *cis*-5'b led to lower yields of *cis*-16'b (Table 4). Finally, acid-catalyzed removal of the protecting groups yielded the arginine dihydrochloride mimetics 17, which were purified by reverse-phase C-18 column chromatography. Under a high concentration of HCl (6 M), partial epimerization occurred at the C-2 position in the case of methyl carbamates 16a. To avoid

Table 5NOS-inhibitory activity of 17

	nNOS IC ₅₀	iNOS IC50	eNOS IC ₅₀
(\pm) -cis-17	>1 mM	80 μM	200 μM
(\pm) -trans-17	>1 mM	160 μM	150 μM

such epimerization, methyl carbamates **16a** were reacted with trimethylsilyl iodide³⁴ instead of aqueous HCl, leading to **17** in approximately 80% conversion, along with approximately 10% of its methylcarbamate with no detectable epimerization.

The inhibition potency of the final arginine mimetics **17** was evaluated by examining their effects on the NOS-dependent oxidation of L-arginine to L-citrulline, according to the procedure previously described by Bredt *et al.*³⁵ The results of inhibition with three NOS isoforms (nNOS, iNOS and eNOS), are summarized in Table 5. Neither of the diastereomers of **17** showed inhibition towards the neuronal NOS. Although weak, some promising effects were observed in the case of inducible and endothelial isoforms. Moreover, some selectivity was observed: *cis*-**17** was more efficient in inhibiting iNOS, and *trans*-**17** was slightly more efficient in the case of the eNOS isoform (Table 5).

Conclusion

In summary, the reactivity of 5,6-dehydropipecolate derivatives **3** towards various electrophilic reagents was examined. This study allowed efficient and diastereoselective preparation of various 5-substituted pipecolates, which are valuable intermediates for the synthesis of potentially bioactive piperidine derivatives. The value of this reactivity study was illustrated by the synthesis of both diastereomers of 5-guanidinopipecolate, which we designed as constrained mimetics of arginine, and whose NOS-inhibitory activity towards three isoforms was evaluated. Weak to moderate inhibition was found and the development of less constrained



Scheme 12 Conversion of azides 5 into 5-guanidinopipecolates 17.

Table 4	Guanylation	conditions	of	amines	15
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Azide	Amine	Guanylation conditions	16 (%)
trans-5a	trans-15a	TfN=C(NHBoc) ₂ , DIPEA	trans-16a (77)
trans-5a	trans-15a	BocN=C(SMe)NHBoc, HgCl ₂ , Et ₃ N	<i>trans</i> -16a (61)
trans-5b	trans-15b	CbzN=C(SMe)NHCbz, Et ₃ N	<i>trans</i> -16b (56)
trans-5b	trans-15b	CbzN=C(SMe)NHCbz, HgCl ₂ , Et ₃ N	<i>trans</i> -16b (80)
cis-5a	cis-15a	TfN=C(NHBoc) ₂ , DIPEA	<i>cis</i> -16a (68)
cis-5a	cis-15a	BocN=C(SMe)NHBoc, HgCl ₂ , Et ₃ N	<i>cis</i> -16a (85)
cis-5b	cis-15b	CbzN=C(SMe)NHCbz, Et ₃ N	<i>cis</i> -16b (52)
cis-5b	cis-15b	CbzN=C(SMe)NHCbz, HgCl ₂ , Et ₃ N	<i>cis</i> -16b (83)
trans-5'b	trans-15'b	CbzN=C(SMe)NHCbz, Et ₃ N	<i>trans</i> -16'b (58)
cis-5′b	<i>cis</i> -15′b	$TfN=C(NHCbz)_2, Et_3N$	<i>cis</i> -16′b (22)

pipecolate-based arginine mimetics is underway. The possibility of accessing these 5-substituted pipecolates efficiently is an important issue, as they can subsequently be included in peptidic structures. We are currently investigating several fundamental aspects of these compounds, such as their impact upon the conformation of peptides and their use as scaffolds³⁶ for the synthesis of small-molecule libraries.

Experimental

General remarks

IR spectra were recorded with a Perkin Elmer Spectrum one FT-IR spectrometer equipped with a MIRacleTM single reflection horizontal ATR unit (zirconium-selenium crystal).¹H NMR spectra were recorded at 250 MHz on a Bruker AM 250 spectrometer, in CDCl₃ at 300 K (unless otherwise indicated); ¹³C NMR spectra were recorded at 63 MHz on the same instrument. Chemical shifts are reported in parts per million (δ in ppm) and are referenced against solvent signals ($\delta_{\rm C}$ 77.16 for chloroform) for ¹³C spectra and solvent residual resonance ($\delta_{\rm H}$ 7.26 for chloroform) for ¹H spectra. Coupling constants J are given in Hz. Multiplicity designation used are: s, d, t, q, dd, and m for singlet, doublet, triplet, quadruplet and double doublet respectively; broad signals are denoted by br. Mass spectra, chemical ionisation (CI) or fast atom bombardment (FAB) were recorded by the "Service de Spectrometrie de Masse" at Paris Descartes University. All reactions were carried out under an argon atmosphere, and were monitored by thin layer chromatography with Merck 60F-254 precoated (0.2 mm) on glass, and by gas chromatography (GC) analysis performed with an HP6890 apparatus equipped with DB1 capillary column (length: 25 m, diameter: 0.32 m) and an HP 3395 integrator. Dichloromethane (DCM) was distilled from CaH₂ under Ar. THF was distilled, under argon, from sodium/benzophenone ketyl radical immediately prior to use. Flash chromatography was performed with Merck kieselgel 60 (0.2–0.5 mm) or Bakerbond C-18 (0.04 mm); the solvent systems are given as v/v.

The effects of compounds 17 on the NOS-dependent oxidation of L-arginine to L-citrulline were determined according to a previously described protocol.³⁵ Briefly, enzymatic reactions were conducted at 37 °C for 5 min in 50 mM HEPES (pH 7.4) containing 5 mM dithiothreitol (DTT), 1 mM NADPH, 1 mM $CaCl_2$, 10 µg mL⁻¹ calmoduline, 20 µM tetrahydrobiopterin (BH₄), $4 \mu M FAD$, $4 \mu M FMN$, $10 \mu M$ (about 500 000 cpm) [2,3,4,5-³H]-Larginine, and increasing concentrations of the tested compounds. Final incubation volumes were 100 µL. The reactions were started by the addition of protein and terminated by the addition of 500 µL cold stop buffer (20 mM sodium acetate pH 5.5, 1 mM L-citrulline, 2 mM EDTA and 0.2 mM EGTA). Samples (500 µL) were applied to columns containing 1 mL of Dowex AG 50W-X8 (Na⁺ form, prepared from the H⁺ form), pre-equilibrated with stop buffer and a total of 1.5 mL of stop buffer was added to eluate [3H]-L-citrulline. Aliquots were then mixed with Pico-Fluor 40 (Packard) and counted on a Packard Tri-Carb 2300 liquid scintillation spectrometer. Control samples without NOS or NADPH were included for background determinations. Incubations in the presence of inducible NOS were performed similarly but CaCl₂ and calmoduline were omitted.

General methods for preparation of azides 5

Method A (from enecarbamates 3, by azidomethoxylation/ reduction sequence). Azidomethoxylation. To a 0.08 M solution of enecarbamate 3a (or 3b) and NaN₃ (1.5 equiv.) in a 4 : 1 mixture of acetone–MeOH was added dropwise a 0.1 M solution of cerium ammonium nitrate (3 equiv.) in acetone at -95 °C. The mixture was stirred at *ca.* -90 °C until total conversion of 1 (as monitored by TLC), then diluted with H₂O and extracted with Et₂O. The combined organic layers were washed with water and brine, dried over MgSO₄ and concentrated *in vacuo*, prior to purification by flash column chromatography, affording adduct 4. Azidomethoxylation of enecarbamate 1c was performed in acetonitrile instead of acetone; the CAN solution was added at 0 °C and the reaction mixture was stirred at r.t. for 16 h.

Reduction. To a solution of the 5-azido-6-methoxy compound **4** in DCM were added $BF_3 \cdot OEt_2$ (1.05 equiv.) and Et_3SiH (1.05 equiv.) at -90 °C. After allowing warming until r.t. for 8 h, the solution was diluted with DCM, and a saturated aqueous solution of NaHCO₃ was added. The aqueous phase was extracted twice with DCM; the combined organic layers were washed with water, dried over MgSO₄, and concentrated *in vacuo*. The final product **5** was purified by flash column chromatography.

Method B (from mesylates 13 or tosylates 14). To the mesylate or tosylate dissolved in DMF, was added NaN₃ (7–8 equiv.) and the mixture was heated at 65–100 °C; then DMF was removed *in vacuo*. Water (50 mL) was added and the mixture was extracted with DCM. The organic extracts were washed with water, dried over MgSO₄ and concentrated *in vacuo*. Purification by flash column chromatography afforded pure azido compound **5**.

(2*S**,5*R**)-Dimethyl 5-azidopiperidine-1,2-dicarboxylate *trans*-5a. Following Method A, from enecarbamate 3a (275 mg, 1.38 mmol), NaN₃ (137 mg, 2.07 mmol) and CAN (2.27 g, 4.14 mmol), at -90 °C for 7 h. After workup, the crude azidomethoxylation compound 5a was submitted to reduction with Et₃SiH (194 µL, 1.21 mmol) and BF₃·OEt₂ (172 µL, 1.21 mmol). Flash column chromatography (cyclohexane–EtOAc 85 : 15) afforded a *trans/cis* (87 : 13) mixture of azido compounds 5a (58%).

Following Method B, from mesylate *cis*-**13a** (380 mg, 1.29 mmol) and NaN₃ (620 mg, 9.54 mmol) in DMF (5 mL). Flash column chromatography (cyclohexane–EtOAc 7 : 3) afforded *trans*-**5a** as a colourless oil (220 mg, 72%).

Following Method B, from tosylate *cis*-**14a** (90 mg, 0.24 mmol) and NaN₃ (120 mg, 1.82 mmol) in DMF (2 mL). Flash column chromatography (cyclohexane–EtOAc 7 : 3) afforded *trans*-**5a** as a colourless oil (51 mg, 86%).

$$\begin{split} R_{\rm f} &= 0.26 \; ({\rm cyclohexane/AcOEt}\; 7:3); \; \delta_{\rm H} \; (250 \; {\rm MHz}, \; {\rm CDCl}_3; \\ {\rm Me}_4{\rm Si})\; 5.05{-}4.94\; (0.6 \; {\rm H}, {\rm m}, 2{\rm -}{\rm H}), 4.90{-}4.75\; (0.4 \; {\rm H}, {\rm m}, 2{\rm -}{\rm H}), 4.35{-} \\ 4.00\; (1\; {\rm H}, {\rm m}, 6{\rm -}{\rm H}), 3.90{-}3.60\; (7\; {\rm H}, {\rm m}, 5{\rm -}{\rm H}, \; {\rm NCO}_2{\rm CH}_3, \; {\rm CO}_2{\rm CH}_3), \\ 3.40{-}3.10\; (1\; {\rm H}, {\rm m}, 6{\rm -}{\rm H}), \; 2.15{-}2.00\; ({\rm m}, \; 2\; {\rm H}, \; 3{\rm -}{\rm H}), \; 1.95{-}1.45\; ({\rm m}, \\ 2\; {\rm H}, \; 4{\rm -}{\rm H}); \; \delta_{\rm C} \; (63\; {\rm MHz}, \; {\rm CDCl}_3)\; 171.4\; ({\rm CO}_2{\rm CH}_3), \; 156.7\; 156.2\; ({\rm NCO}_2{\rm CH}_3), \; 54.8\; ({\rm CH}{-}5),\; 53.7\; 53.4\; ({\rm CH}{-}2),\; 52.7\; ({\rm NCO}_2{\rm CH}_3), \\ 52.2\; ({\rm CO}_2{\rm CH}_3), \; 44.1\; ({\rm CH}_2{-}6),\; 24.5\; ({\rm CH}_2{-}4),\; 20.8\; ({\rm CH}_2{-}3);\; {\rm MS}\; ({\rm ESI}):\; m/z\; {\rm calcd}\; {\rm for}\; \\ {\rm C_9H_{14}N_4O_4Na\; [{\rm M}+Na]^+\; 265.0913,\; {\rm found\; 265.0910}. \end{split}$$

(2 S^* ,5 S^*)-Dimethyl 5-azidopiperidine-1,2-dicarboxylate *cis*-5a. Following Method B, from mesylate *trans*-13a (380 mg, 1.29 mmol) and NaN₃ (620 mg, 9.54 mmol) in DMF (5 mL). Flash column chromatography (cyclohexane–EtOAc 7 : 3) afforded *cis*-**5a** as a colourless oil (220 mg, 72%). $R_{\rm f} = 0.30$ (cyclohexane–EtOAc 65 : 35); $\delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 4.98–4.85 (0.6 H, m, 2-H), 4.83–4.70 (0.4 H, m, 2-H), 4.40–4.25 (0.4 H, m, 6-H), 4.23–4.05 (0.6 H, m, 6-H), 3.85–3.60 (6 H, m, CO₂CH₃), 3.50–3.20 (1 H, m, 5-H), 2.95–2.65 (1 H, s, 6-H), 2.45–2.20 (1 H, m, 3-H), 2.15–1.95 (1 H, m, 4-H), 1.85–1.60 (1 H, m, 3-H), 1.45–1.15 (1 H, m, 4-H); $\delta_{\rm c}$ (63 MHz, CDCl₃) 171.2 (CO₂CH₃), 156.5, 156.1 (NCO₂CH₃), 56.4 (CH-5), 53.6, 53.3, 52.6 (CH-2, NCO₂CH₃, CO₂CH₃), 45.5, 45.3 (CH₂-6), 26.9 (CH₂-4), 25.5, 25.2 (CH₂-3); MS (ESI): m/z = 265 [M + Na]⁺ 100%; HRMS (ESI) m/z calcd for C₉H₁₄N₄O₄Na [M + Na]⁺ 265.0913, found 265.0924.

(2*S**,5*R**)-1-*tert*-Butyl 2-methyl 5-azidopiperidine-1,2-dicarboxylate *trans*-5b. Following Method A, from enecarbamate 3b (332 mg, 1.38 mmol), NaN₃ (137 mg, 2.07 mmol) and CAN (2.27 g, 4.14 mmol), at -90 °C for 1 h. Reduction step was performed by using BF₃·OEt₂ (172 µL, 1.21 mmol) and Et₃SiH (194 µL, 1.21 mmol). Flash column chromatography (cyclohexane–EtOAc 85 : 15) afforded a *trans/cis* (92 : 8) mixture of azido compound 5b (60%) from which isomer *trans* could be isolated as pure compound (155 mg, 47%).

Following Method B, from mesylate *cis*-**13b** (292 mg, 0.87 mmol) and NaN₃ (450 mg, 6.92 mmol) in DMF (3.5 mL). Flash column chromatography (cyclohexane–EtOAc 85 : 15) afforded pure *trans*-**5b** as a colourless oil (225 mg, 91%).

$$\begin{split} R_{\rm f} &= 0.23 \; ({\rm cyclohexane-EtOAc} \; 8:\; 2); \; v_{\rm max}({\rm neat})/{\rm cm}^{-1} \; 2981, \\ 2945, 2108, 1726, 1679, 1416, 1364, 1243, 1147, 1013; &_{\rm H}\; (250\;{\rm MHz}, \\ {\rm CDCl}_{\rm s}; \; {\rm Me}_4 {\rm Si})\; 5.05-4.85\; (0.6\;{\rm H}, {\rm m}, 2-{\rm H}), \; 4.83-4.60\; (0.4\;{\rm H}, {\rm m}, 2-{\rm H}), \\ 4.25-4.00\; (1\;{\rm H}, {\rm m}, 6-{\rm H}), \; 3.85-3.60\; (4\;{\rm H}, {\rm m}, 5-{\rm H}, {\rm CO}_2{\rm CH}_3), \\ 3.35-3.00\; (1\;{\rm H}, {\rm m}, 6-{\rm H}), \; 2.10-1.95\; (2\;{\rm H}, {\rm m}, 3-{\rm H}), \; 1.90-1.30\; \\ (11\;{\rm H}, {\rm m}, 4-{\rm H}, {\rm CMe}_3); \; \delta_{\rm C}\; (63\;{\rm MHz}, {\rm CDCl}_3)\; 171.9\; ({\rm CO}_2{\rm CH}_3), \\ 155.3\; ({\rm CO}_2 t{\rm Bu}), \; 80.7\; ({\rm CMe}_3), \; 55.0\; ({\rm CH}{-5}), \; 54.2, \; 52.9\; ({\rm CH}{-2}), \\ 52.2\; ({\rm CO}_2 {\rm CH}_3), \; 44.2, \; 43.7\; ({\rm CH}_2{-6}), \; 28.2\; ({\rm C}Me_3), \; 24.9\; ({\rm CH}_2{-4}), \\ 21.0\; ({\rm CH}_2{-3}); \; {\rm MS}\; ({\rm ESI}): \; m/z\; = \; 307\; [{\rm M} + {\rm Na}]^+\; 100\%; \; {\rm HRMS}\; \\ ({\rm ESI})\; m/z\; {\rm calcd}\; {\rm for}\; {\rm C}_{12}{\rm H}_{20}{\rm N}_4{\rm O}_4{\rm Na}\; [{\rm M} + {\rm Na}]^+\; 307.1382,\; {\rm found}\; 307.1374. \end{split}$$

 $(2S^*,5S^*)$ -1-*tert*-Butyl 2-methyl 5-azidopiperidine-1,2-dicarboxylate *cis*-5b. Following method B, from mesylate *trans*-13b (1.48 g, 4.39 mmol) and NaN₃ (2.28 g, 35.12 mmol) in DMF (16 mL). Flash column chromatography (cyclohexane–EtOAc 85 : 15) afforded pure *cis*-5b as a colourless oil (1.11 g, 90%).

$$\begin{split} R_{\rm f} &= 0.32 \; ({\rm cyclohexane-EtOAc}\;8\,:\,2);\; \nu_{\rm max}({\rm neat})/{\rm cm}^{-1}\;2971,\\ 2868,\;2098,\;1741,\;1695,\;1403,\;1364,\;1245,\;1147;\;\delta_{\rm H}\;(250\;{\rm MHz},\\ {\rm CDCl}_3;\;{\rm Me}_4{\rm Si})\;4.95-4.80\;(0.6\;{\rm H},\,{\rm m},\;2-{\rm H}),\;4.75-4.60\;(0.4\;{\rm H},\,{\rm m},\\ 2-{\rm H}),\;4.35-4.17\;(0.4\;{\rm H},\,{\rm m},\;6-{\rm H}),\;4.25-3.98\;(0.6\;{\rm H},\,{\rm m},\;6-{\rm H}),\;3.72\;\\ (3\;{\rm H},\,{\rm s},\;{\rm CO}_2{\rm CH}_3),\;3.45-3.20\;(1\;{\rm H},\,{\rm m},\;5-{\rm H}),\;2.85-2.55\;(1\;{\rm H},\,{\rm m},\\ 6-{\rm H}),\;2.45-2.20\;(1\;{\rm H},\,{\rm m},\;3-{\rm H}),\;2.10-1.90\;(1\;{\rm H},\,{\rm m},\;4-{\rm H}),\;1.85-1.60\;\\ (1\;{\rm H},\,{\rm m},\;3-{\rm H}),\;1.55-1.15\;(10\;{\rm H},\,{\rm m},\;4-{\rm H},\;{\rm CMe}_3);\;\delta_{\rm C}\;(63\;{\rm MHz},\\ {\rm CDCl}_3\;)\;171.3\;({\rm CO}_2{\rm CH}_3),\;155.1,\;154.8\;({\rm CO}_2{-}t-{\rm Bu}),\;80.8\;({\rm CMe}_3),\\ 56.4\;({\rm CH}{-}5),\;53.8,\;52.5\;({\rm CH}{-}2),\;52.2\;({\rm CO}_2{\rm CH}_3),\;45.7,\;44.6\;({\rm CH}_2{-}6),\\ 28.2\;({\rm CM}e_3),\;26.8\;({\rm CH}_2{-}4),\;25.2,\;25.0\;({\rm CH}_2{-}3);\;{\rm MS}\;({\rm ESI}):\;m/z=\\ 307\;[{\rm M}+{\rm Na}]^+\;100\%;\;{\rm HRMS}\;({\rm ESI})\;m/z\;{\rm calcd}\;{\rm for}\;{\rm C}_{12}{\rm H}_{20}{\rm N}_4{\rm O}_4{\rm Na}\\[{\rm M}+{\rm Na}]^+\;307.1382,\;{\rm found}\;307.1367.\\ \end{split}$$

General method for hydroboration/oxidation sequences

Hydroboration. To a solution of the enecarbamate **3** in DCM at -80 °C was added dropwise a 2 M solution of BH₃·SMe₂ (1 equiv.)

in THF, and the mixture then warmed to room temperature. The reaction mixture was stirred at r.t. for 2.5 h (unless otherwise indicated), and then concentrated *in vacuo* to afford a borane compound, which was used in the oxidative steps.

Oxidation with Me_3NO (for preparation of alcohols 8). To the crude hydroboration compound were added THF and trimethylamine *N*-oxide and the heterogeneous mixture was refluxed at 70 °C for 15 min then concentrated *in vacuo*. Water was added and the organic layer was extracted with DCM, dried over MgSO₄, and then concentrated *in vacuo*. Purification by flash column chromatography afforded alcohols 8.

Oxidation with iodoxy benzoic acid (IBX) (for preparation of ketones 12). To the crude hydroboration compound were added acetonitrile and IBX (4 equiv.) and the heterogeneous mixture was refluxed at 80 °C for 2.5 h. After filtration over Celite and washing with DCM, the filtrate was concentrated *in vacuo*. The residue was diluted with H₂O and the organic layer was extracted with DCM. Purification by flash column chromatography afforded ketones 12.

(2S*, 5R*) - Dimethyl 5 - hydroxypiperidine - 1, 2 - dicarboxylate *trans*-8a^{23b}. Prepared according to the above procedure, starting from enecarbamate 3a (2.01 g, 10.1 mmol) in DCM (30 mL) and BH₃·SMe₂ (2 M in THF, 5.0 mL, 10.1 mmol); then oxidation with triethylamine N-oxide (6.7 g, 60.6 mmol) in THF (30 mL). Flash column chromatography (EtOAc-cyclohexane 7 : 3) afforded a trans/cis (85:15) mixture of alcohol 8a (63%) from which pure sample of the trans isomer could be isolated as a colourless oil (1.1 g, 50%): $R_{\rm f} = 0.20$ (EtOAc-cyclohexane 7 : 3); $v_{\rm max}({\rm neat})/{\rm cm}^{-1}$ $3451, 3016, 2955, 1738, 1686, 1447, 1252, 1123, 1017; \delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 4.98 (0.7 H, br s, 2-H), 4.84 (0.3 H, br s, 2-H), 4.15-3.85 (2 H, m, 6-H, 5-H), 3.73 (6 H, br s, CO₂CH₃, NCO₂CH₃), 3.30-3.13 (1 H, m, 6-H), 2.25-1.95 (2 H, m, 4-H), 1.85-1.40 (2 H, m, 3-H); $\delta_{\rm C}$ (63 MHz, CDCl₃) 171.9 (CO₂CH₃), 157.5 (NCO₂CH₃), 63.4 (CH-5), 54.2, 53.8 (CH-2), 53.0, 52.3 (CO₂CH₃), 47.5, 42.6 (CH_2-6) , 27.1, 26.7 (CH_2-4) , 20.2 (CH_2-3) ; MS (ESI): m/z = 235 $[M + NH_4]^+$ 100%.

(2S*,5R*)-1-tert-Butyl 2-methyl 5-hydroxypiperidine-1,2-dicarboxylate trans-8b. Prepared according to the above procedure, starting from enecarbamate 3b (5 g, 20.75 mmol) in DCM (100 mL), and BH3 ·SMe2 (2 M in THF, 10.4 mL, 20.80 mmol), stirring at r.t. was maintained for 4 h, prior to oxidation with triethylamine N-oxide (12.5 g, 112.6 mmol) in THF (100 mL). Flash column chromatography (EtOAc-cyclohexane 6 : 4) afforded a trans/cis (93:7) mixture of alcohol 8b (3.4 g, 63%) as colourless oil: $R_{\rm f} = 0.23$ (EtOAc-cyclohexane 6 : 4); $\delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 5.05-4.65 (1 H, m, 2-H), 4.15-3.80 (2 H, m, 5-H, 6-H), 3.70 (3 H, s, CO₂CH₃), 3.30–2.95 (1 H, m, 6-H), 2.30–1.60 (4 H, m, 3-H, 4-H), 1.42 (9 H, s, t-Bu); $\delta_{\rm C}$ (63 MHz, CDCl₃) 172.5 (CO₂CH₃), 156.6 (CO₂-t-Bu), 80.8 (CMe₃), 63.9 (CH-5), 54.9, 53.7 (CH-2), 52.4 (CO₂CH₃), 48.2, 47.3 (CH₂-6), 28.6 (CMe₃), 27.3 (CH₂-4), 20.6 (CH₂-3); MS (ESI): m/z = 282 [M + Na]⁺ 100%; HRMS (ESI) m/z calcd for C₁₂H₂₁NO₅Na [M + Na]⁺ 282.1317, found 282.1312.

 $(2S^*,5S^*)$ -Dimethyl 5-hydroxypiperidine-1,2-dicarboxylate *cis*-8a^{23b}. This stereoisomer was isolated in low yield from the oxidation-hydroboration of enecarbamate 3a. It was also prepared, with higher yield, by reduction of ketone 12a (2.58 g,

12.0 mmol) in ethanol (45 mL) by portionwise addition of sodium borohydride (910 mg, 23.9 mmol) at 0 °C, then the mixture was stirred at 0 °C for 15 min. The reaction mixture was portionned between DCM (50 mL) and 10% aqueous solution of citric acid (pH = 6). The aqueous layer was re-extracted with EtOAc $(2 \times 30 \text{ mL})$. The organic extracts were combined, dried over MgSO₄ and concentrated in vacuo. Purification by flash column chromatography (EtOAc) afforded a cis/trans (99:1) mixture of alcohol 8a (1.9 g, 74%): $R_{\rm f} = 0.26$ (EtOAc-cyclohexane 7 : 3); $v_{\rm max}$ (neat)/cm⁻¹ 3418, 2953, 2866, 1736, 1682, 1446, 1237, 1212, 1151, 998; $\delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 4.86 (0.6 H, br s, 2-H), 4.85 (0.4 H, br s, 2-H), 4.24 (0.4 H, d, J 11.0, 6-H), 4.12 (0.6 H, d, J 11.0, 6-H), 3.75–3.53 (7H, m, CO₂CH₃, NCO₂CH₃, 5-H), 2.83-2.64 (1 H, m, 6-H), 2.31-2.23 (1 H, m, 3-H), 2.00-1.91 (1 H, m, 4-H), 1.80–1.63 (1 H, m, 3-H), 1.31–1.10 (1 H, m, 4-H); δ_c (63 MHz, CDCl₃) 171.5 (CO₂CH₃), 157.3 (NCO₂CH₃), 66.5 (CH-5), 53.5, 53.2 (CH-2), 53.1, 52.4 (CO₂CH₃), 48.0 (CH₂-6), 30.3 (CH₂-4), 24.9 (CH₂-3); MS (ESI): $m/z = 235 [M + NH_4]^+$ 25%.

(2S*,5S*)-1-tert-Butyl 2-methyl-cis-5-hydroxypiperidine-1,2dicarboxylate cis-8b. Obtained in higher quantities by reduction of ketone 12b (832 mg, 3.24 mmol) in MeOH (17 mL), with sodium borohydride (129 mg 3.39 mmol) at 0 °C following the procedure employed in the case of ketone 12a. Purification by flash column chromatography (EtOAc-cyclohexane 55:45) afforded a cis/trans (97:3) mixture of alcohol 8b (712 mg, 85%) from which pure cis-**8b** could be isolated as colourless oil (630 mg, 75%): $R_{\rm f} = 0.21$ (EtOAc-cyclohexane 55 : 45); $v_{max}(neat)/cm^{-1}$ 3416, 2971, 2868, 1739, 1692, 1403, 1238, 1209, 1142, 997; $\delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 4.90-4.55 (1 H, m, 2-H), 4.30-3.95 (1 H, m, 6-H), 3.80-3.50 (4 H, m, CO₂CH₃, 5-H), 2.85–2.55 (1 H, m, 6-H), 2.35–2.15 (1 H, m, 3-H), 2.05–1.60 (3 H, m, 3-H, 4-H, OH), 1.43 (9 H, s, t-Bu), 1.30–1.10 (1 H, m, 4-H); $\delta_{\rm C}$ (63 MHz, CDCl₃) 170.4 (CO₂CH₃), 155.5, 155.2 (CO2-t-Bu), 80.6 (CMe3), 66.4, 66.3 (CH-5), 54.0, 52.7 (CH-2), 52.2 (CO₂CH₃), 48.4, 47.6 (CH₂-6), 30.4, 29.8 (CH₂-4), 28.3 (CMe₃), 25.1, 24.9 (CH₂-3); MS (ESI): m/z = 282 [M + Na]⁺ 100%; HRMS (ESI) m/z calcd for C₁₂H₂₁NO₅Na [M + Na]⁺ 282.1317, found 282.1327.

rac-Dimethyl 5-oxopiperidine-1,2-dicarboxylate 12a^{29a}. Hydroboration of enecarbamate 3a(1.1 g, 5.53 mmol) with BH₃·SMe₂ (2 M in THF, 2.76 mL, 5.52 mmol) in DCM (17 mL) then oxidative treatment with IBX (6.19 g, 22.1 mmol) in CH₃CN (75 mL), according to the procedure described above. Flash column chromatography (EtOAc-cyclohexane 5 : 5) afforded ketone 12a as a yellow oil (600 mg, 51%): $R_{\rm f} = 0.34$ (EtOAccyclohexane 6 : 4); v_{max} (neat)/cm⁻¹ 2958, 1737, 1694, 1441, 1203; $\delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 4.85 (0.6 H, t, J 6.1, 2-H), 4.71 (0.4 H, t, J 6.1, 2-H), 4.46 (0.4 H, d, J 19.0, 6-H), 4.32 (0.6 H, d, J 19.0, 6-H), 3.93 (0.6 H, d, J 19.0, 6-H), 3.88 (0.4 H, d, J 19.0, 6-H), 3.76–3.72 (6 H, m, CO₂CH₃), 2.51–2.10 (4 H, m, 3-H, 4-H); δ_c (63 MHz, CDCl₃) 204.6 (C-5), 171.8 (CO₂CH₃), 156.3 (NCO₂CH₃), 53.7, 53.4 (CH-2), 53.2, 52.5 (CO₂CH₃), 51.9, 51.5 (CH₂-6), 35.6 (CH₂-4), 23.9, 23.5 (CH₂-3); MS (ESI): m/z = 233 $[M + NH_4]^+ 100\%$.

rac-1-tert-Butyl **2-methyl 5-oxopiperidine-1,2-dicarboxylate 12b**^{29b}. Hydroboration of enecarbamate **3b** (438 mg, 1.82 mmol) with $BH_3 \cdot SMe_2$ (2 M in THF, 0.91 mL, 1.82 mmol) in DCM

(9 mL), at r.t. for 4 h; and subsequent oxidative treatment with IBX (2.04 g, 7.82 mmol) in CH₃CN (10 mL), according to the procedure described above. Flash column chromatography (cyclohexane–EtOAc 7 : 3) afforded ketone **12b** as a yellow oil (187 mg, 40%): $R_f = 0.24$ (cyclohexane–EtOAc 7 : 3); $v_{max}(neat)/cm^{-1}$ 2971, 1736, 1695, 1392, 1150; δ_H (250 MHz, CDCl₃; Me₄Si) 4.85–4.75 (0.6 H, m, 2-H), 4.65–4.50 (0.4 H, m, 2-H), 4.38 (0.6 H, d, *J* 19.0, 6-H), 4.27 (0.4 H, d, *J* 19.0, 6-H), 3.97–3.80 (1 H, m, 6-H), 3.75 (3 H, s, CO₂CH₃), 2.50–1.95 (4 H, m, 3-H, 4-H), 1.43 (9 H, m, *t*-Bu); δ_C (63 MHz, CDCl₃) 205.6 (C-5), 172.6, 172.3 (CO₂CH₃), 154.9, 154.4 (CO₂-*t*-Bu), 81.4 (CMe₃), 54.6, 53.1 (CH-2), 52.5 (CO₂CH₃), 51.0 (CH₂-6), 36.0, 35.8 (CH₂-4), 28.3 (CMe₃), 23.8, 23.7 (CH₂-3); MS (ESI): m/z = 256 [M – H]⁺ 100%.

(2*S**,5*R**)-Dimethyl *trans*-5-(*N*,*N*'-bis(*tert*-butoxycarbonyl)-guanidino)piperidine-1,2-dicarboxylate *trans*-16a

Method A. To a solution of the amino ester *trans*-15a (0.73 g, 2.89 mmol) in DCM (40 mL), were added N,N'-bis(*tert*-butoxycarbonyl)-N''-trifluoromethanesulfonylguanidine (3 g, 7.67 mmol) and DIPEA (1.41 mL, 8.69 mmol). The heterogeneous mixture was stirred at r.t. for 5 days and then concentrated *in vacuo*. Purification by flash column chromatography (DCM \rightarrow DCM–MeOH 98 : 2) afforded the 5-guanidino compound *trans*-16a as a white solid (1.02 g, 77%).

Method B. To a solution of the amino ester trans-15a (103 mg, 0.41 mmol) in DCM (5 mL) was added N,N'-bis(tertbutoxycarbonyl)-S-methylisothiourea (277 mg, 0.96 mmol), triethylamine (157 µL, 1.13 mmol) and HgCl₂ (265 mg, 0.98 mmol). The mixture was stirred at r.t. for 30 h, then filtered through a pad of Celite, washed with DCM and the filtrate was concentrated in vacuo. Flash column chromatography (DCM-DCM-MeOH 98 : 2) afforded the 5-guanidino compound trans-16a as a white solid (115 mg, 61%). v_{max} (neat)/cm⁻¹ 2975, 1740, 1714, 1633, 1613, 1564, 1442, 1413, 1321, 1246, 1148; $R_{\rm f} = 0.42$ (DCM/CH₃OH 98 : 2); m.p. = 64–68°C; $\delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 11.42 (s, 1 H, NHBoc), 8.80 (1 H, br s, NH), 5.05-4.70 (1 H, m, 2-H), 4.55-4.26 (1 H, m, 5-H), 4.25–3.95 (1 H, m, 6-H), 3.73 (3 H, s, CO₂CH₃), 3.71 (3 H, s, NCO₂CH₃), 3.30–3.15 (1 H, m, 5-H), 2.22–2.07 (1 H, m, 3-H), 2.01–1.77 (2 H, m, 4-H), 1.51–1.44 (19 H, m, 2 × CMe₃, 3-H); $\delta_{\rm C}$ (63 MHz, CDCl₃) 171.4 (CO₂CH₃), 163.6, 155.5, 153.1 (N-C=O, C=N), 83.2, 79.2 (2×CMe₃), 53.8 (CH-2), 53.0, 52.5 $(CO_2CH_3, NCO_2CH_3), 45.7 (CH_2-6), 43.7 (CH-5) 28.3, 28.0 (2 \times$ CMe_3), 24.9 (CH₂-4), 21.3 (CH₂-3); MS (ESI): m/z = 481 [M + Na]⁺ 100%; HRMS (ESI) m/z calcd for C₂₀H₃₄N₄O₈Na [M + Na]⁺ 481.2274, found 481.2260.

(2*S**,5*S**)-Dimethyl *trans*-5-(*N*,*N*'-bis(*tert*-butoxycarbonyl)guanidino)piperidine-1,2-dicarboxylate *cis*-16a

The amino ester *cis*-**15a** (580 mg, 2.29 mmol) was guanylated with N,N'-bis(*tert*-butoxycarbonyl)-*S*-methylisothiourea following the procedure described above for its *trans* isomer. Flash column chromatography (DCM \rightarrow DCM–MeOH 98 : 2) afforded the 5-guanidino compound *cis*-**16a** as a white solid (898 mg, 85%). $v_{\text{max}}(\text{neat})/\text{cm}^{-1}$ 2979, 1715, 1637, 1611, 1560,1446, 1411, 1368, 1335, 1309, 1242, 1137, 1052; δ_{H} (250 MHz, CDCl₃; Me₄Si) 11.48 (1 H, br s, NHBoc), 8.80 (1 H, d, *J* 7.8, *N*H), 4.98–4.88 (0.6 H,

m, 2-H), 4.82–4.70 (0.4 H, m, 2-H), 4.40–3.98 (2 H, m, 5-H, 6-H), 3.80–3.60 (6 H, m, CO₂CH₃), NCO₂CH₃), 2.85–2.60 (1 H, m, 6-H), 2.35–2.18 (1 H, m, 3-H), 2.10–1.92 (1 H, m, 4-H), 1.91–1.69 (1 H, m, 3-H), 1.65–1.35 (18 H, m, 2 × CMe₃), 1.30–1.05 (1 H, m, 4-H); $\delta_{\rm C}$ (63 MHz, CDCl₃) 171.80 (CO₂CH₃), 163.8, 156.8, 156.5, 155.9, 153.4 (N-C=O, C=N), 83.6; 79.8, 79.6 (CMe₃), 53.7, 53.6, 53.3, 53.2, 52.6 (CH-2, CO₂CH₃), NCO₂CH₃), 46.1 (CH-5), 45.8 (CH₂-6) 28.4, 28.2 (CMe₃), 27.7, 27.6 (CH₂-3), 25.7, 25.4 (CH₂-4); MS (ESI): m/z = 481 [M + Na]⁺ 100%; HRMS (ESI) m/z calcd for C₂₀H₃₄N₄O₈Na [M + Na]⁺ 481.2274, found 481.2256.

(2*S**,5*R**)-1-*tert*-Butyl 2-methyl-5-(*N*,*N*'-bis(benzyloxycarbonyl)-guanidino)piperidine-1,2-dicarboxylate *trans*-16b

To a solution of the amino ester trans-15b (80.5 mg, 0.33 mmol) in DCM (4 mL), were added N,N'-bis(benzyloxycarbonyl)-Smethylisothiourea (328 mg, 0.92 mmol), triethylamine (55 μ L, 0.39 mmol) and mercury dichloride (255 mg, 0.94 mmol). The mixture was stirred at r.t. for 16 h then concentrated in vacuo. Purification by flash column chromatography (DCM \rightarrow DCM– MeOH 97 : 3) afforded the 5-guanidino compound trans-16b as a colourless oil (150 mg, 80%). $R_{\rm f} = 0.46$ (DCM–MeOH 99 : 1); $v_{\rm max}$ (neat)/cm⁻¹ 3322, 3291, 2950, 1798, 1728, 1695, 1633, 1617, 1566, 1452, 1423, 1377, 1364, 1336, 1300, 1240, 1199, 1124, 1046; $\delta_{\rm H}$ (250 MHz, CDCl₃; Me₄Si) 11.67 (1 H, s, NHZ), 8.79 (1 H, d, J 7.0, NH), 7.44–7.08 (10 H, m, H_{AL}), 5.20–4.65 (5 H, m, 2 × OCH₂Ph, 2-H), 4.40–3.95 (2 H, m, 5-H, 6-H), 3.68 (3 H, s, CO₂CH₃), 3.22-2.93 (1 H, m, 6-H), 2.20-2.00 (1 H, m, 3-H), 1.98-1.72 (2 H, m, 3-H, 4-H), 1.63–1.16 (10 H, m, CMe₃, 4-H); $\delta_{\rm C}$ (63 MHz, CDCl₃) 171.7 (CO₂CH₃), 163.8, 155.7, 155.4, 153.9 (N-C=O, C=N), 136.8, 134.7 (C_{Ar}), 128.9, 128.7, 128.6, 128.5, 128.2, 128.0 (CH_{Ar}), 80.9 (CMe₃), 68.3, 67.3 ($2 \times OCH_2Ph$), 54.6, 53.1 (CH-2), 52.4 (CO₂CH₃), 45.5 (CH₂-6), 44.8, 44.5 (CH-5), 28.3 (CMe_3) , 24.9 (CH_2-4) , 21.4 (CH_2-3) ; MS (ESI): m/z = 591 [M + Na]⁺ 100%; HRMS (ESI) m/z calcd for C₂₉H₃₆N₄O₈Na [M + Na]⁺ 591.2431, found 591.2402.

(2*S**,5*S**)-1-*tert*-Butyl 2-methyl -5-(*N*,*N*'-bis-(benzyloxycarbonyl)guanidino)piperidine-1,2- dicarboxylate *cis*-16b

Guanvlation of the cis-amino ester compound cis-15b (93 mg, 0.38 mmol) was carried out as for its trans isomer. Flash column chromatography (DCM \rightarrow DCM–MeOH 97 : 3) afforded the 5guanidino compound *cis*-16b as a colourless oil (181 mg, 83%). $R_{\rm f} = 0.44 \,({\rm DCM-MeOH}\,99:1); v_{\rm max}({\rm neat})/{\rm cm}^{-1}\,3271,2976,1785,$ 1741, 1728, 1690, 1646, 1620, 1584, 1452, 1431, 1382, 1367, 1333, 1297, 1281, 1263, 1212, 1147, 1052; δ_H (250 MHz, CDCl₃; Me₄Si) 11.73 (1 H, s, NHZ), 8.16 (1 H, br s, NH), 7.46-7.17 (10 H, m, H_{Ar}), 5.25–5.00 (4 H, m, 2 × OC H_2 Ph), 4.95–4.62 (1 H, m, 2-H), 4.35–3.93 (2 H, m, 5-H, 6-H), 3.72 (3 H, s, CO₂CH₃), 2.70–2.53 (1 H, m, 6-H), 2.35-2.13 (1 H, m, 3-H), 2.06-1.90 (1 H, m, 4-H), 1.88–1.65 (1 H, m, 3-H), 1.41 (9 H, s, CMe₃), 1.28–1.05 (1 H, m, 4-H); $\delta_{\rm C}$ (63 MHz, CDCl₃) 171.8 (CO₂CH₃), 163.8, 155.6, 155.3, 154.9, 153.9 (N-C=O, C=N), 136.7, 134.6 (C_{Ar}), 128.9, 128.8, 128.5, 128.4, 128.2, 128.0 (CH_{Ar}), 80.7 (CMe₃), 68.3, 67.3 $(2 \times OCH_2Ph)$, 53.9, 52.6 (CH-2), 52.3 (CO₂CH₃), 46.7 (CH-5), 45.7, 44.9 (CH2-6), 28.3 (CMe3), 27.6 (CH2-4), 25.2 (CH2-3); MS (ESI): $m/z = 591 [M + Na]^+ 100\%$; HRMS (ESI) m/z calcd for $C_{29}H_{36}N_4O_8Na [M + Na]^+ 591.2431$, found 591.2435.

5-guanidinopiperidine-2-carboxylic acids 17

Method A. Guanidino compound **16** (0.5 mmol scale) was heated at reflux in 6 M aqueous HCl (6 mL) for 4 days then concentrated *in vacuo*. The resulting powder was submitted to purification by C-18 reverse phase column chromatography (H₂O) to afford the final arginine mimetic **17** (69–79%).

Method B (for derivatives 16a). Guanidino compound 16a (0.1-0.3 mmol scale) was refluxed in chloroform (1-3 mL) in the presence of trimethylsilyl iodide (16 equiv.) for 24 h. The reaction mixture was quenched by addition of 3 mL of saturated methanol with HCl(g) then concentrated under vacuum prior to purification by C-18 reverse phase column chromatography (H₂O) as in Method A.

 $\begin{array}{l} \textbf{(25*,5R*)-17 (trans isomer).} \quad \delta_{\rm H} \ (250 \ {\rm MHz}, \ {\rm D_2O}) \ 3.95-3.77 \\ (2 \ {\rm H}, \ {\rm m}, \ 2-{\rm H}, \ 5-{\rm H}), \ 3.76-3.62 \ (1 \ {\rm H}, \ {\rm m}, \ 6-{\rm H}), \ 3.15-2.95 \ (1 \ {\rm H}, \ {\rm m}, \ 6-{\rm H}), \ 2.58-2.41 \ (1 \ {\rm H}, \ {\rm m}, \ 3-{\rm H}), \ 2.39-2.25 \ (1 \ {\rm H}, \ {\rm m}, \ 4-{\rm H}), \ 2.05-1.65 \ (2 \ {\rm H}, \ {\rm m}, \ 3-{\rm H}), \ 4-{\rm H}); \ \delta_{\rm C} \ (63 \ {\rm MHz}, \ {\rm D_2O}) \ 174.2 \ ({\rm CO_2H}), \ 159.1 \\ ({\rm C=N}), \ 59.4 \ ({\rm CH-2}), \ 48.4 \ ({\rm CH_2-6}), \ 48.1 \ ({\rm C-5}) \ 31.1 \ ({\rm CH_2-4}), \ 27.2 \\ ({\rm CH_2-3}); \ {\rm MS} \ ({\rm FAB}): \ m/z = 187 \ [{\rm M} + {\rm H}]^+ \ 100\%; \ {\rm Anal. \ Calcd. \ for} \\ {\rm C_7H_{16}Cl_2N_4O_2 \cdot 0.75H_2O}: \ {\rm C}, \ 30.84; \ {\rm H}, \ 6.47; \ {\rm N}, \ 20.55. \ {\rm Found: \ C}, \ 30.94; \ {\rm H}, \ 6.39; \ {\rm N}, \ 20.13. \end{array}$

 $\begin{array}{l} \textbf{(2S*,5S*)-17 (cis isomer).} \quad \delta_{\rm H} \ (250 \ {\rm MHz}, D_2 {\rm O}) \ 4.17-3.97 \ (2 \ {\rm H}, \\ {\rm m}, \ 2-{\rm H}, \ 5-{\rm H}), \ 3.58-3.39 \ (2 \ {\rm H}, \ {\rm m}, \ 6-{\rm H}), \ 2.34-2.06 \ (3 \ {\rm H}, \ {\rm m}, \ 3-{\rm H}), \\ 4-{\rm H}), \ 2.02-1.82 \ (1 \ {\rm H}, \ {\rm m}, \ 4-{\rm H}); \ \delta_{\rm C} \ (63 \ {\rm MHz}, \ D_2 {\rm O}) \ 173.8 \ ({\rm CO}_2 {\rm H}), \\ 159.2 \ ({\rm C=N}), \ 58.3 \ ({\rm CH-2}), \ 47.7 \ ({\rm CH}_2-6), \ 46.9 \ ({\rm CH-5}), \ 28.4 \ ({\rm CH}_2-4), \\ 24.3 \ ({\rm CH}_2-3); \ {\rm MS} \ ({\rm FAB}): \ m/z \ = \ 187 \ [{\rm M} \ + \ {\rm H}]^+ \ 100\%; \ {\rm Anal}. \\ {\rm Calcd.} \ {\rm for} \ C_7 {\rm H}_{16} {\rm Cl}_2 {\rm N}_4 {\rm O}_2 \cdot 1.75 {\rm H}_2 {\rm O}: \ {\rm C}, \ 28.93; \ {\rm H}, \ 6.76; \ {\rm N}, \ 19.28. \\ {\rm Found:} \ {\rm C}, \ 28.39; \ {\rm H}, \ 6.40; \ {\rm N}, \ 19.81. \end{array}$

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